

## Datasheet: MMT1

<b>Description:</b>	MOUSE MONOCLONAL ANTIBODY ISOTYPING TEST KIT
<b>Name:</b>	MOUSE ISOTYPING KIT
<b>Format:</b>	Kit
<b>Product Type:</b>	Kits
<b>Quantity:</b>	10 TESTS

## Product Details

### Applications

This product has been reported to work in the following applications. This information is derived from testing within our laboratories, peer-reviewed publications or personal communications from the originators. Please refer to references indicated for further information. For general protocol recommendations, please visit [www.bio-rad-antibodies.com/protocols](http://www.bio-rad-antibodies.com/protocols).

	Yes	No	Not Determined	Suggested Dilution
Isotyping Assay	■			

Where this product has not been tested for use in a particular technique this does not necessarily exclude its use in such procedures. Suggested working dilutions are given as a guide only. It is recommended that the user titrates the product for use in their own system using appropriate negative/positive controls.

### Target Species

Mouse

### Product Information

The mouse monoclonal antibody isotyping test kit shows no cross-reactivity with bovine IgG (<0.1 %).

### Test Principle

The assay principle is based on anti-mouse kappa and anti-mouse lambda antibodies coupled onto coloured micro particles and equally reactive to any mouse monoclonal antibody regardless of its isotype. The isotyping strip has immobilized bands of goat anti-mouse antibodies corresponding to each of the common mouse antibody isotypes (IgG1, IgG2a, IgG2b, IgG3, IgM, and IgA) and to the kappa and lambda light chains. Both sides of the strip bear a positive flow control band, which indicates that the antibody-coated coloured micro particles have migrated through the strip. By using these two components, a mouse monoclonal antibody can be screened for isotype by simply diluting the antibody sample, pipetting the diluted sample into the development tube where it forms a complex with the antibody coated micro particles, and inserting the strip. This complex flows through the strip until it is bound by the immobilized goat anti-mouse antibody specific for the monoclonal's isotype and its light chain. In approximately 5-10 minutes, the micro particle complex will aggregate as blue bands in the two sections corresponding to the monoclonal antibody's isotype and its light chain. Development of the strip is complete when the positive flow control band on each side of the test strip turns blue. Please note, this kit is not suitable for use with antibodies of the isotype IgG2c; some cross-reactivity may be observed with isotype IgG2a.

### Reagents In The Kit

1 Desiccant vial containing mouse isotyping test strips, 10 tests.  
10 Capped ready-to-use lyophilized microparticle development tubes.

### Instructions For Use

**Note:** All reagents should be brought to room temperature before use.

### Sample Preparation:

**Dilute all monoclonal antibody samples to a concentration of 1.0 ug/ml in PBS containing 1% w/v bovine serum albumin (BSA).** If the concentration of the sample is entirely unknown, make dilutions based on the following estimates:

Typically, serum contains between 10-15 mg/ml IgG and ascites can be as high as 10 mg/ml. Hollow fibre bioreactor culture supernatants contain approximately 0.5-1.0 mg/ml, whereas static flask tissue culture supernatants usually contain 10-50 ug/ml. Using these estimates, the appropriate dilutions can be made.

150ul of the diluted sample will be added to the development tubes.

#### **Assay Protocol:**

1. Remove the required number of isotyping strips from the desiccant vial and replace the cap. Remove the caps from an equal number of development tubes.

**Note:** the tubes may be labeled with a marker for identification.

2. Pipette 150ul of the freshly diluted sample into each development tube and incubate at room temperature for 30 seconds. Vortex the tube briefly to ensure that the coloured micro particle solution is completely re-suspended.

3. Place one isotyping strip, with the solid red end at the bottom, into each development tube.

#### **Interpretation of Results:**

Interpret the results at 5-10 minutes once the positive flow control bands have appeared. Within 5-10 minutes, a blue band will appear above the letters in one of the class or subclass windows as well as in either the kappa or lambda window of the strip, indicating the heavy and light-chain composition of the monoclonal antibody. The intensity of the blue bands will increase as the sample continues to flow up the strip. The positive flow control bands on each side of the isotyping test strip should also appear, indicating that the antibody-coated micro particles are functional and have flowed up the strip. In cases where the sample is very dilute, the development time may take up to 10 minutes.

**Note:** For a permanent experimental record or for an easier interpretation of results when testing multiple samples, the solid red area may be cut off the bottom of the strip to prevent further band development once the positive flow control bands have appeared. A gentle stream of air can be applied to the membrane portion of the strip to assist in drying the membrane and preventing any further development. **Do not wash the strip to stop the reaction.**

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#### **References**

1. Kimura, H. *et al.* (2008) The organization of histone H3 modifications as revealed by a panel of specific monoclonal antibodies. [Cell Struct Funct. 33: 61-73.](#)
2. Gilbert, J.A. *et al.* (2006) Monoclonal pathogenic antibodies to the thyroid-stimulating hormone receptor in Graves' disease with potent thyroid-stimulating activity but differential blocking activity activate multiple signaling pathways. [J Immunol. 176 \(8\): 5084-92.](#)
3. Keim, S.A. *et al.* (2008) Generation and characterization of monoclonal antibodies against the proregion of human desmoglein-2. [Hybridoma \(Larchmt\). 2008 Aug;27\(4\): 249-58.](#)
4. Stenman, U.H. *et al.* (2011) Elimination of complement interference in immunoassay of hyperglycosylated human chorionic gonadotropin. [Clin Chem. 57: 1075-7.](#)
5. Hayashi-Takanaka, Y. *et al.* (2011) Tracking epigenetic histone modifications in single cells using Fab-based live endogenous modification labeling. [Nucleic Acids Res. 39: 6475-6488.](#)
6. Nakayama, E. *et al.* (2011) Antibody-dependent enhancement of Marburg virus infection. [J Infect Dis. 204 Suppl 3: S978-85.](#)

7. Urán, M.E. *et al.* (2011) Detection of antibodies against *Paracoccidioides brasiliensis* melanin in *in vitro* and *in vivo* studies during infection. [Clin Vaccine Immunol. 18: 1680-8.](#)
8. Hayashi-Takanaka, Y. *et al.* (2015) Distribution of histone H4 modifications as revealed by a panel of specific monoclonal antibodies. [Chromosome Res. Sep 5. \[Epub ahead of print\]](#)
9. Álvarez, B. *et al.* (2015) Molecular and functional characterization of porcine Siglec-3/CD33 and analysis of its expression in blood and tissues. [Dev Comp Immunol. 51 \(2\): 238-50.](#)
10. Leng J *et al.* (2014) Development of a novel anti ESAT-6 monoclonal antibody for screening of *Mycobacterium tuberculosis*. [Int J Clin Exp Med. 7 \(11\): 4238-43.](#)
11. Broecker, F. *et al.* (2016) Multivalent display of minimal *Clostridium difficile* glycan epitopes mimics antigenic properties of larger glycans. [Nat Commun. 7: 11224.](#)
12. Arimitsu, H. *et al.* (2016) Immunochromatographic detection of the heat-labile enterotoxin of enterotoxigenic *Escherichia coli* with cross-detection of cholera toxin [Journal of Microbiological Methods. Dec 8 \[Epub ahead of print\]](#)
13. Álvarez, B. *et al.* (2015) Molecular and functional characterization of porcine Siglec-3/CD33 and analysis of its expression in blood and tissues. [Dev Comp Immunol. 51 \(2\): 238-50.](#)
14. Okda, F. *et al.* (2015) Development of an indirect ELISA, blocking ELISA, fluorescent microsphere immunoassay and fluorescent focus neutralization assay for serologic evaluation of exposure to North American strains of Porcine Epidemic Diarrhea Virus. [BMC Vet Res. 11: 180.](#)

<b>Storage</b>	Store at +4°C. DO NOT FREEZE. Do not use components from different lots.
<b>Shelf Life</b>	Please see label for expiry date.
<b>Health And Safety Information</b>	Material Safety Datasheet documentation available at: Not required
<b>Regulatory</b>	For research purposes only
<b>Technical Advice</b>	<u>PROBLEM</u>

**No heavy and or light-chain band appeared on the strip, but the positive flow control bands appeared.**

*Possible causes:*

1. The antibody concentration was too low - prepare a less dilute sample and re-test.
2. No antibody was in the sample - the hybridoma is either not secreting or is not a mouse monoclonal. If possible sub-clone the hybridoma and re-test.
3. Freshly diluted samples were not used - prepare fresh dilutions and re-test.

PROBLEM

**Multiple heavy and light-chain bands appear on the strip.**

*Possible causes:*

1. Antibody concentration was too high - dilute sample further and re-test.
2. For ascites, there may be small amounts of contaminating antibodies produced - dilute sample further and re-test.
3. For tissue culture supernatant, a mixed culture may be present - re-clone the hybridoma and re-test.

## PROBLEM

**No positive flow control bands appear.**

*Possible causes:*

1. Sample volume was too low (<150ul) - carefully dilute a fresh sample and pipette 150ul into a new development tube and re-test.
2. Strip removed from development tube too early - re-test and allow strip to react for at least 10 minutes.

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